

Management of insect vectored diseases

2024

Growing Wisconsin Conference

(Formally known as Wisconsin fresh Fruit and Vegetable Conference)

Harvesting Knowledge, Cultivating connections,
Producing quality and Promoting success



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Presentation Outline

- Fundamentals of pathogen spread by insects (definitions)
 - association with insects (circulative, propagative)
 - transmission types (non-persistent, semi-persistent, persistent)
 - patterns of spread (primary vs secondary)
 - insect movement and colonization

- Managing the problem (approach)
 - varietal selection
 - sanitation and inoculum source reduction
 - precise timing of crop protectants



Photo credit. Gerald Holmes

Insect - Vectored, Plant Disease Ecology

Spread of Plant Pathogens by Insects Involves Interactions of

Plant

Pathogen

Insect Vector

Environment

weather

cultural practices

farming operations

agroecosystem



Plant
Disease



whiteflies



thrips



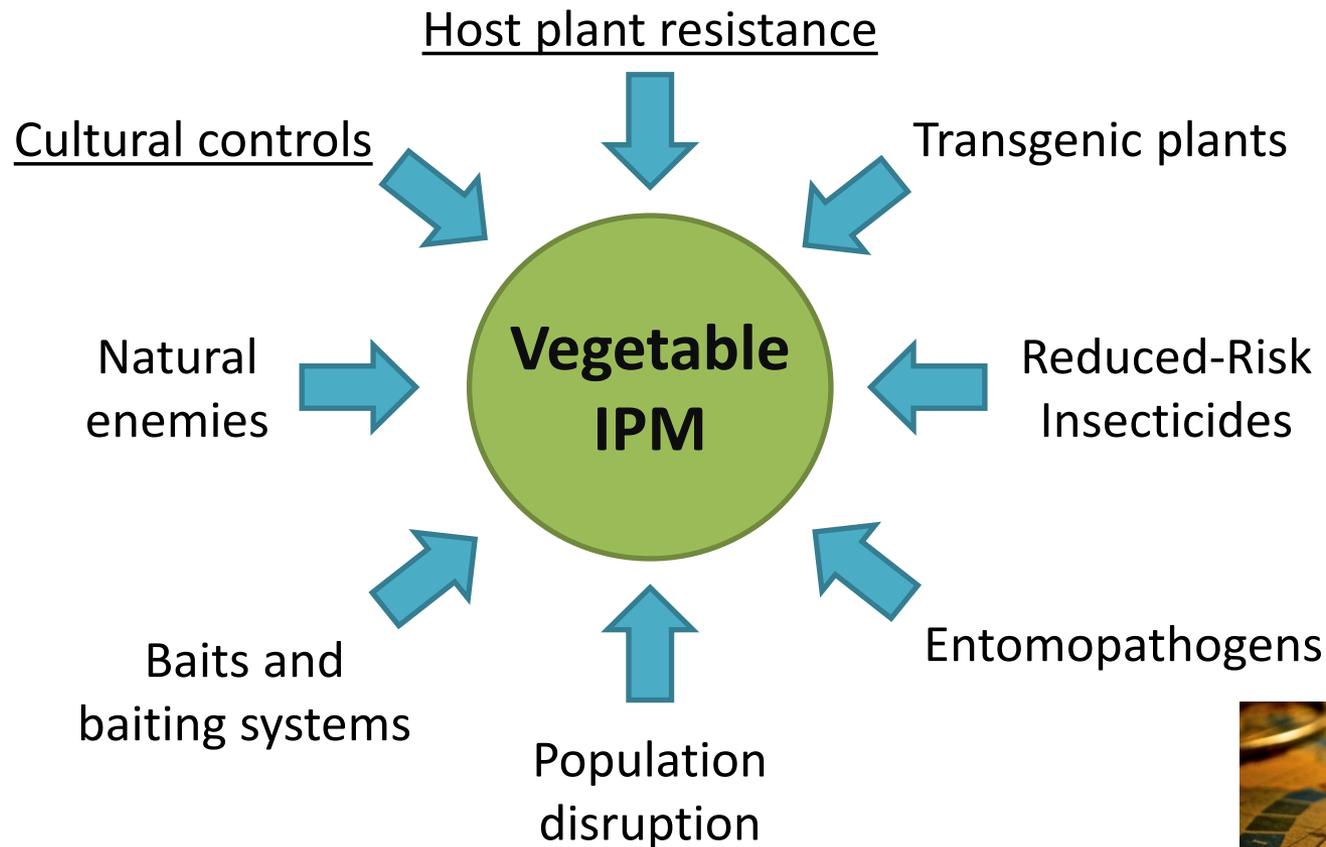
aphids



leafhoppers

Pest management tactics for vegetable insects

Integration of available tools to manage pest damage in the most economically, socially, and environmentally sound way



Several non-persistently transmitted viruses

- Cucumber mosaic virus (CMV) – Bromoviridae (Cucumovirus)
- Watermelon mosaic virus (WMV-1) – Potyviridae (Potyvirus)
- Papaya ringspot virus (PRSV-W) – Potyviridae (Potyvirus)
 - Symptoms similar across many viruses
 - Time of infection important
 - Aphid vectors
 - Mixed infections

- Squash mosaic virus (SqMV) – Secoviridae (Comovirus)
 - Beetle transmitted (striped / spotted cucumber beetle)

CMV – Cucumber mosaic virus



Photo credit. Gerald Holmes



Watermelon mosaic virus - WMV



Papaya ringspot virus – PRSV-W



Papaya ringspot virus

Squash mosaic virus - SqMV



Bacterial wilt - *Erwinia tracheiphila*



Striped cucumber beetle



Aster yellows- *Candidatus asteris phytoplasma*



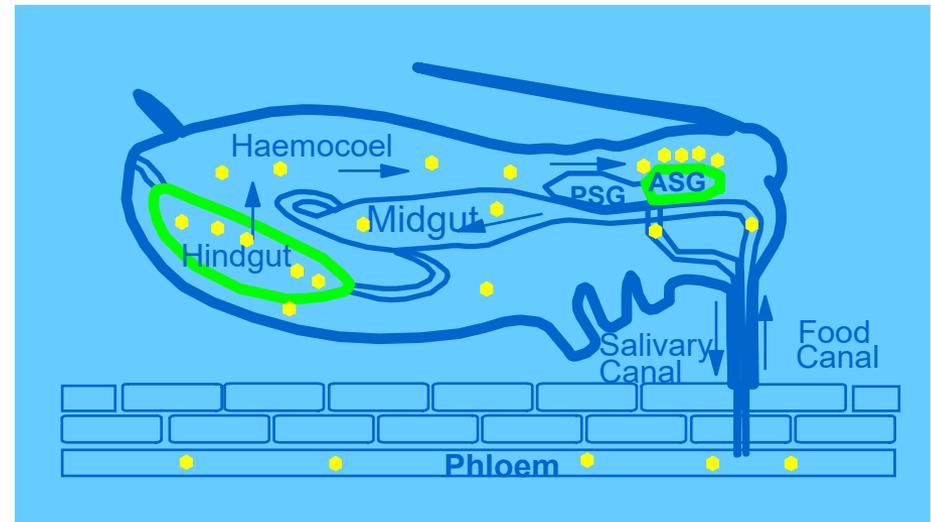
Aster leafhopper



Types of Pathogen Interactions with Insects

Circulative (PLRV, AYp, TSWV)

Systemic movement within insect body

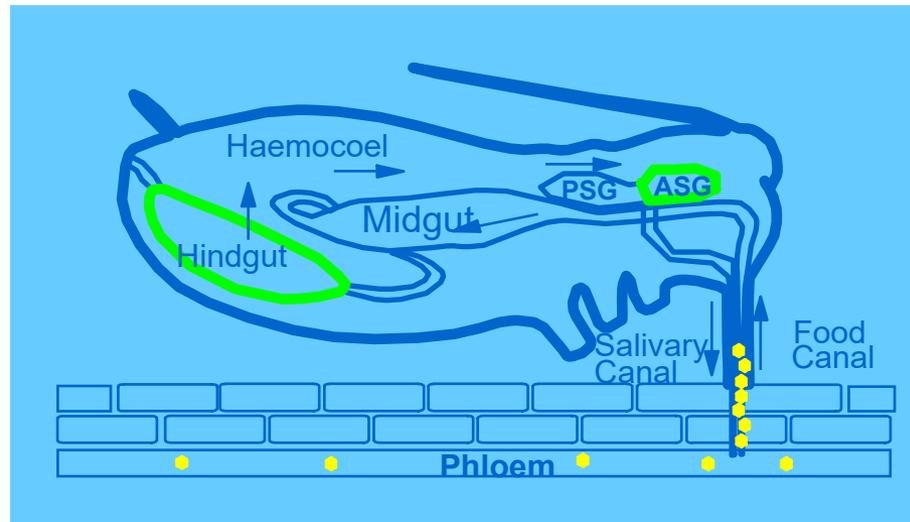


Propagative

Pathogen replication within insect body (transovariol)

Types of Pathogen Interactions with Insects

Non-circulative (PVY, CMV, AMV)
often referred to as “stylet-borne”

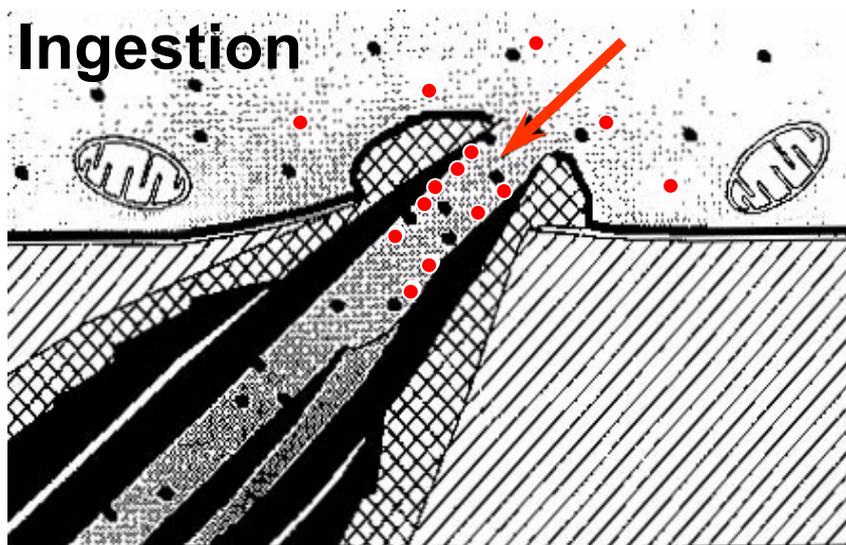


Non-Circulative Pathogen Movement in Insects

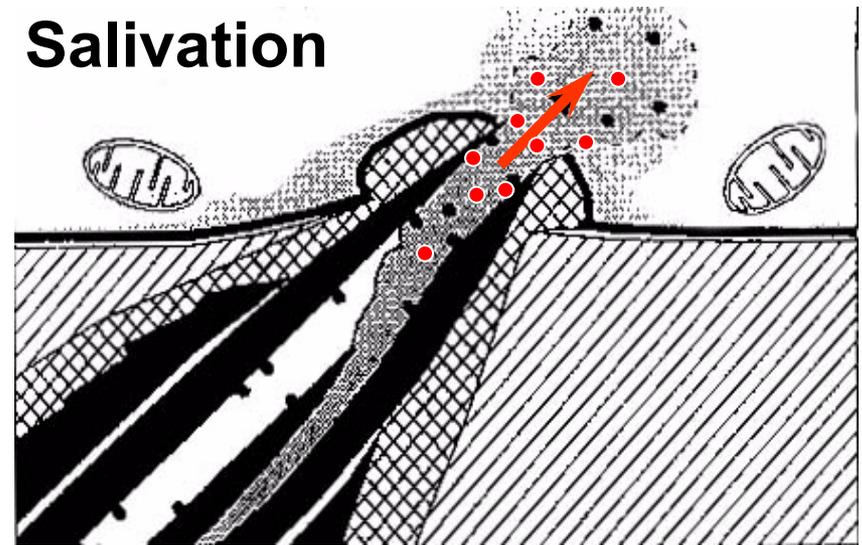
Food Ingestion - pathogen particles attach to maxillary lumen

Egestion - pathogen particles released with saliva

Ingestion



Salivation



Types of Pathogen Transmission by Insects

Non-persistent

often referred to as “stylet-borne”



Persistent

Variations include circulative, propagative, transovariol

Semi-persistent

Categories of Vector Residence

Transient vectors

- do not colonize the crop to which they spread virus
- most important for non-persistent viruses
- relatively unimportant for persistent viruses



Rhopalosiphum padi

Resident vectors

- colonize the crop to which they spread virus
- can be important for all types of transmission
- most important for persistent viruses



Myzus persicae

Traps for Determining Transient Vectors



- Resident vectors are evaluated through visual inspection and associated offspring

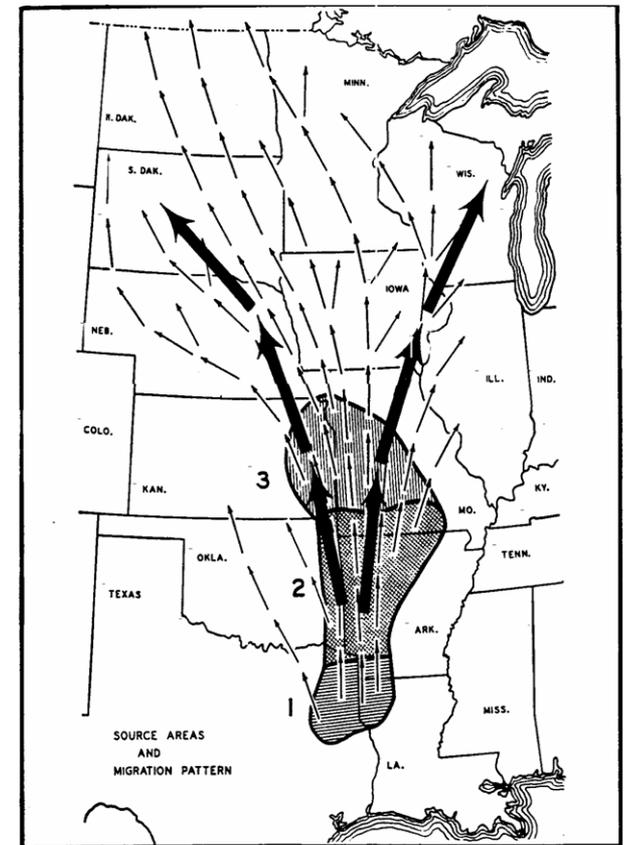
Categories of Insect Dispersal

Trivial flights (aphids, cucumber beetles)

- random movement among selected hosts
- important for secondary spread
- important for non-persistent viruses

Insect migration (aster leafhopper)

- long distance obligatory flights
- can be important for all types of transmission



Host selection and alightment by aphids

- Migrating aphids ready to land are attracted to yellow/green and repelled by UV
- Landing on plants is usually edge-distributed, but generally random
- Suitability of plant determined by briefly probing the plant with mouth parts, or stylets (tasting)
 - Small proportion of aphids remain on 1st suitable host plant encountered
 - Subsequent flights of aphids that leave a suitable plant are relatively short 'trivial'

Reflective Mulch for Aphid/Virus Management

Theory:

Repels winged aphids

(reflected UV)

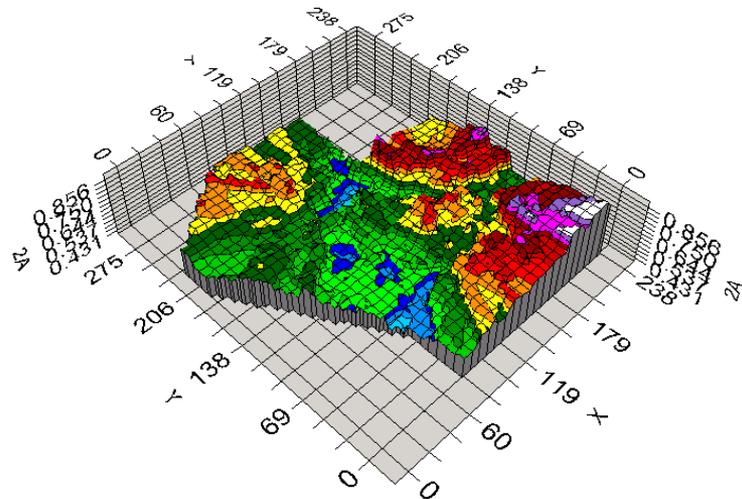
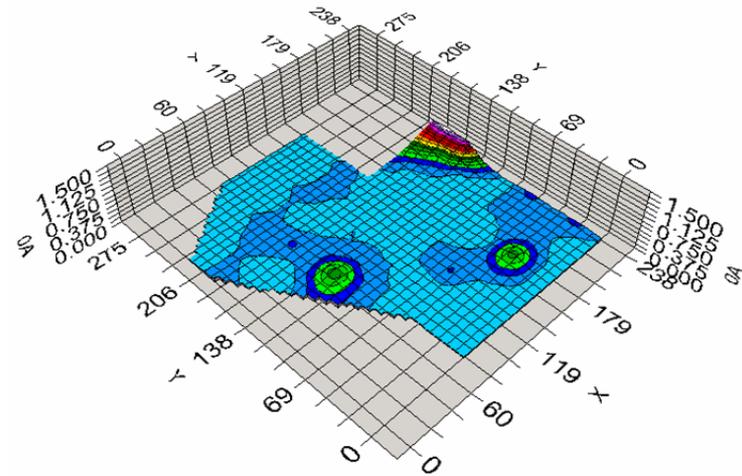
Delays aphid colonization

Delays virus infection



Categories of Pathogen Spread

- **Primary spread:**
 - Initial spread of pathogen into a field from sources outside field
- **Secondary spread:**
 - Spread of pathogen within a field from sources of virus within the same field



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Photo credit. Gerald Holmes

Key Determinants of Virus Spread

- Amount of virus inoculum in area (initial inoculum)
- Location of virus inoculum relative to crop (outside/inside)
- Susceptibility of crop to virus infection (cultivar variation)
- Vectors
 - number of species (relative abundance in landscape)
 - transmission efficiency of vectors
 - abundance of individuals
 - activity of vector
 - level and timing
- Environmental conditions (wind, RH, temp)

General Approaches to Management of Aphid-borne Plant Viruses

- Host manipulation (isolation in space and time), host tolerance (planting dates, variety selection)
- Reducing virus sources (inoculum id and reduction)
- Manipulating (controlling) vectors (alightment, landing, plant apparency)
- Blocking virus transfer between aphid & plant (oils, insecticides, transgenics)

Resistance to virus and bacterial infection



Cornell Disease Resistant Cultivars

Pumpkin Variety	Cucumber Mosaic Virus	Cucumber Vein Yellowing Virus	Papaya Ringspot Virus (WMV-1)	Watermelon Mosaic Virus (Strain 2)	Zucchini Yellow Mosaic Virus	Seed Company
	CMV	CVYV	PRSV	WMV	ZYMV	
Casperita F1 (white mini)				X		HO, HS, JO, SI, SW
Corvette F1				X	X	HO
Harvest Moon					X	C
Jamboree F1	X		X			HO
Magician F1					X	C, HS, RS, SW
Moonstone					X	C
Munchkin (mini)				X	X	HS
Newton F1				X		HO
Poco Blanco (mini)				X		C
Rembrandt F1				X		HO
Silver Moon F1 (white)					X	HO, HS, SI
Speckled Hound F1					X	HS, SI, SW
Speckled Pup F1					X	HS
Zeus F1					X	HS, RS, SW

<http://vegetablemndonline.ppath.cornell.edu/Tables/TableList.htm>

Insect - Sampling and thresholds (foliar)

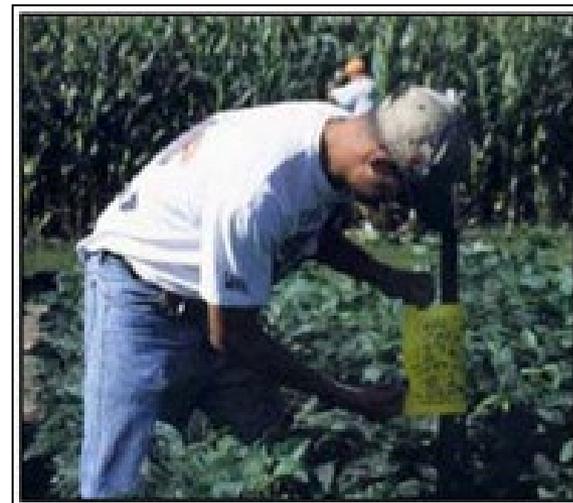
Sampling

- Yellow sticky cards
- Plant counts
- Colonization times critical



Action thresholds

- 1 beetle/plant for melons, cucumbers, and young pumpkins
- 5 beetles/plant for watermelon, squash, and older pumpkins



Reducing sources of inoculum

Weed management



Cover cropping / no till

Sanitation – weed management and exclusion

Cultural

- Eliminate weeds, weedy edges (non-crop sanitation)
- Crop rotation
- Early season row cover

Row cover



Aphid lifecycles

- Two-host species (3 host species)
- Polyphagous

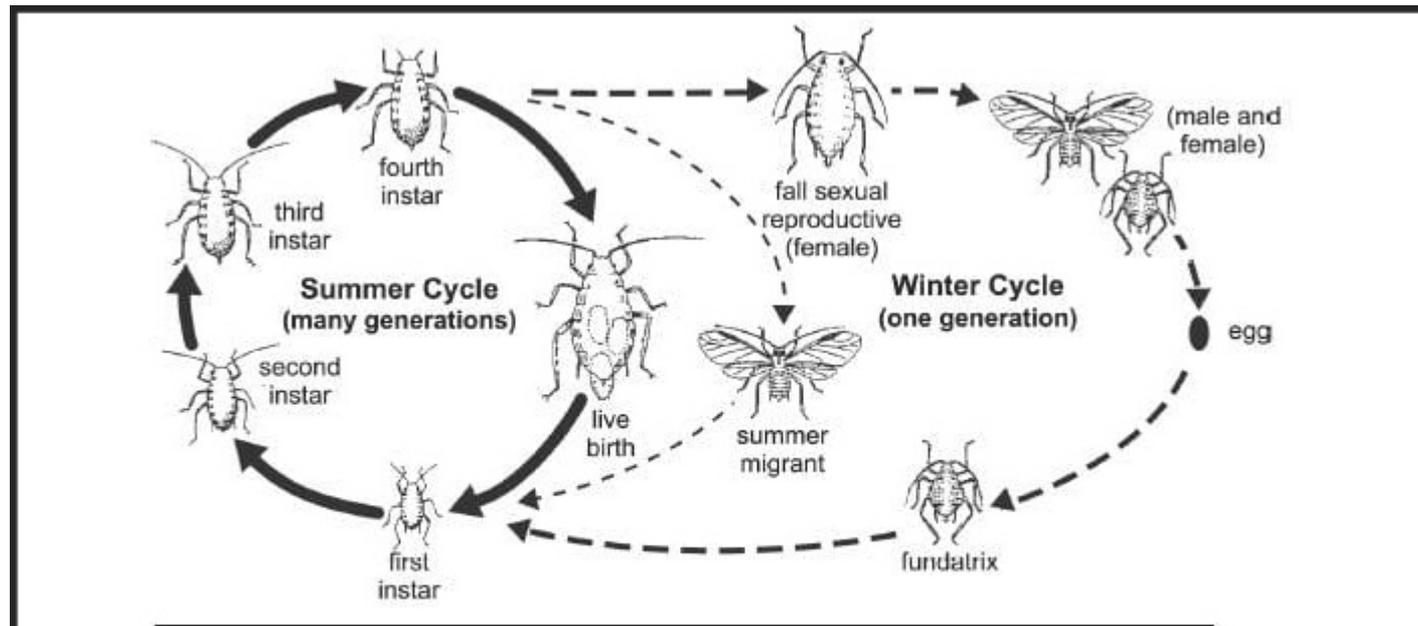


Figure 2. General life cycle of aphids. Asexual reproduction occurs during most of the year (summer cycle). Some aphid species produce a generation of sexual individuals that produce overwintering eggs as shown in the winter cycle.

Aphid generation time(s) and arrival

- Parthenogenic populations (asexual reproduction)
- Temperature dependent generation times
 - *Aphis craccivora* (4.7 days) cowpea aphid
 - *Aphis gossypii* (5.0 days) melon aphid
 - *Rhopalosiphum padi* (5.1 days) bird cherry-oat aphid
 - *Rhopalosiphum maidis* (5.0 days) corn leaf aphid
 - *Aphis nasturtii* (7.8 days) buckthorn aphid
 - *Myzus persicae* (9.1 days) green peach aphid
 - *Macrosiphum euphorbiae* (10.3 days) potato aphid
- Arrival in crop by winged aphids (alatae)
- Colonization in crop typified by non-winged aphids (apterous)
- Initial populations found around edges
 - Pivot (road)
 - Irrigation stands (tracks)
 - Field edges
 - Wind breaks (eddy effects)

Scouting to identify colonizing aphids



- Scout 10-12 sites per field
- Turn over 25 whole leaves per site
- Turn leaves from mid- to lower canopy positions
- Thresholds:
 - '0' alatae (winged)
 - Avg 5 apterae (wingless) per leaf (action threshold)
 - > 15% of leaves infested (4/25)

2020 Applied Research Highlights

University of Wisconsin-Madison
Vegetable Crop Entomology
Extension and Research

Wisconsin Vegetable Entomology

Research ▾ Extension ▾ Crops & Insects ▾ IPM ▾ Lab Members 🔍

Field Trials

Annual reports for summer field research improved pest management recommendations and are available below from 2002 to present. Tabular summaries of experiments performed are shown for recent years. [Browse all trials on our Box server.](#)

2023 Field Trials

Crop	Location	Description	Target Pest(s)	Appl.	Trts	Evaluation	Report
Potato	HARS	Full-Season CPB Trial	Colorado potato beetle	At-plant; foliar	16	Insect counts; yield	PDF
Potato	HARS	Systemic Diamide Trial	Colorado potato beetle (1st gen.), colonizing aphids, potato leafhopper	At-plant; hill side-dress	9	Insect counts	PDF
Potato	HARS	CPB Foliar Trial	Colorado potato beetle (1st gen.)	Foliar	24	Insect counts	PDF
Potato	HARS	Single-app CPB Foliar Trial	Colorado potato beetle (1st gen.)	Foliar	9	Insect counts	PDF
Potato	HARS	Calantha CPB Foliar Timing Trial	Colorado potato beetle (1st gen.)	Foliar	5	Insect counts	PDF
Potato	HARS	Calantha Tank Mix Effects CPB Foliar	Colorado potato beetle (1st gen.)	Foliar	17	Insect counts	PDF

2023 Applied Research Highlights

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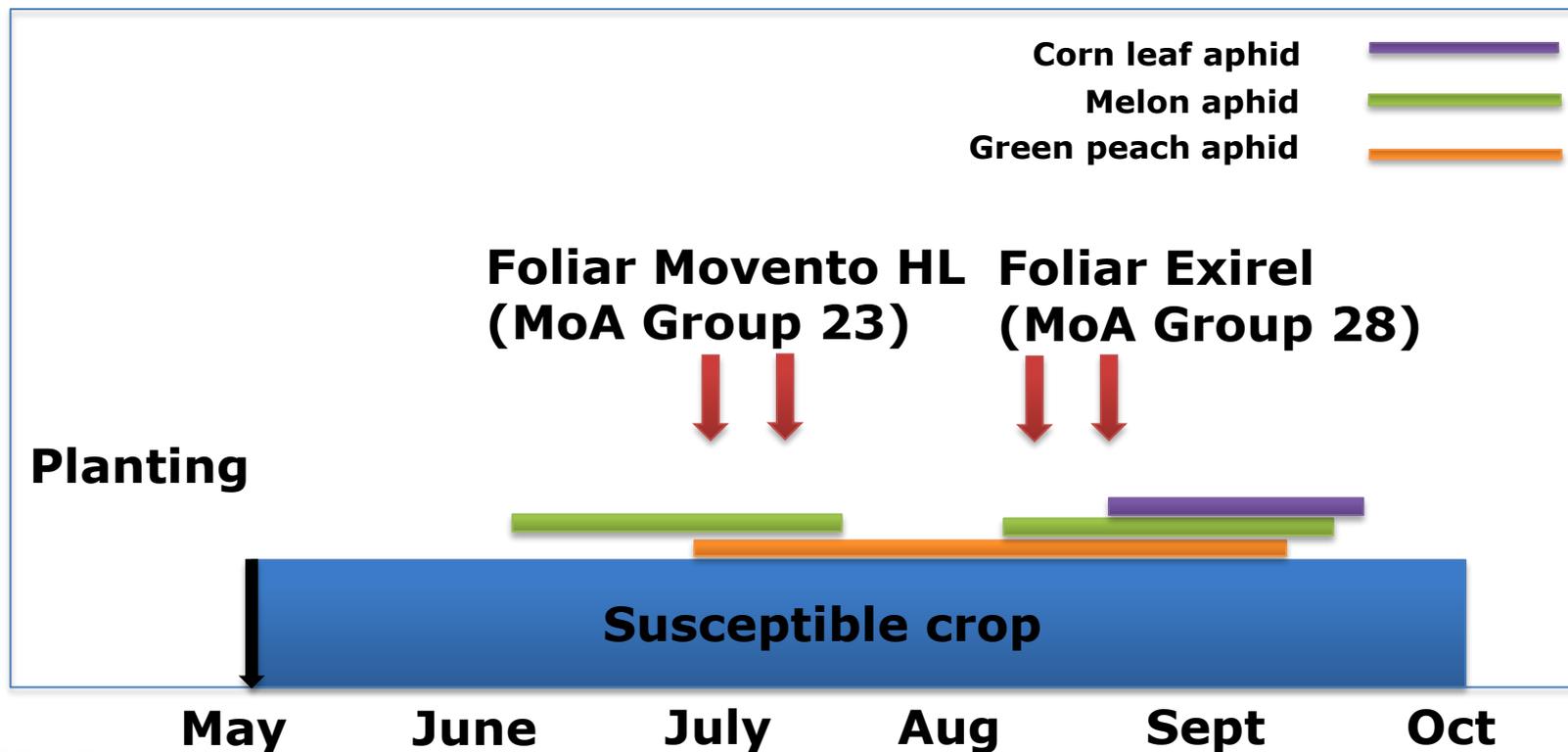
New, recent or existing registrations

- **PQZ** (pyrifluquinazon, Nichino America) – brassicas, cucurbits, potato, leafy: aphids only (Group 9B)
- **Sefina/Versys** (afidopyropen, BASF) – brassicas, leafy, potato: aphids only (Group 9D)
- **Sivanto HL** (flupyradifurone, Bayer Crop Sci) – beans, peas, sweet corn, brassicas, potato: aphids, PLH (Group 4D)(soil and foliar)
- **PFR-97** (*Isaria fumosorosea* Apopka Strain 97, Certis USA) – aphids only; (Group UNK, biologic)
- **Transform WG** (sulfoxaflor, Corteva) – aphids and PLH
beans, peas, sweet corn, brassicas, carrot, onion, potato:
aphids, PLH (Group 4C)



Managing Aphids: Multi-tactic Approach

- Colonizing aphids and mass flights (late June – early July)
- Choice of product should consider other pests



Management considerations

- Populations can build quickly under warm and dry conditions
- Synthetic pyrethroids can 'flare' colonizing aphid populations
- Application coverage is critical to target colonizing species in mid- to low canopy
- Reduced-risk or soft chemistry can conserve beneficial insects that parasitize aphids



Virus Management Summary

- Approaches to limit aphid/beetle/leafhopper landing / alightment
- Selection of cultivars with stated resistance to virus infection
- Sanitation of weedy hosts supporting both the vector (aphids/leafhoppers) and the virus (weedy inoculum)
- Improved understanding of the timing of disease spread and relationship to regional, primary insect vectors – *A. glycines*, *R. padi*, etc...
- Appropriate timing or applications of mineral oil, insecticides or feeding deterrents.

Acknowledgements and Questions



<http://labs.russell.wisc.edu/vegento/>